Chemistry of Phosphorus Ylides. Part 22 [1]. Effect of Newly Synthesized Niclosamide Mannich Bases, Phosphopyranones, Phosphoranylidenes, and Oxaphosphinin on Some Metabolic Aspects of Biomphalaria Alexandrina

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Summary. Niclosamide reacts with secondary amines and formaldehyde under the condition of Mannich reaction, to give new Mannich bases. The reaction of these niclosamide Mannich bases with active phosphacumulene ylides affords the corresponding phenyliminopyranone, pyranone, and pyranthione, respectively. When Wittig reaction was carried out on the pyranone, using p-nitrobenzaldehyde, a new arylidene and triphenylphosphine oxide were obtained. On the other hand, stabilized phosphonium ylides affect the transylidation of niclosamide Mannich bases to the corresponding phosphoranylidenes. When diphenylmethylenetriphenylphosphorane reacts with a niclosamide Mannich base, an oxaphosphinin was obtained. The molluscicidal potency of the newly synthesized derivatives against Biomphalaria alexandrina was studied, too.

Keywords. Niclosamide; Phosphonium ylides; Pyranone derivatives; Transylidation; Phosphoranylidenes; Oxaphosphinin; Molluscicidal potency.

Introduction

Niclosamide [5-chloro-N-(2-chloro-4-nitrophenyl)-2-hydroxybenzamide] (1) is the active ingredient of bayluscide, which has been used as molluscicide of great significance in the last decade [2]. It is widely used in control programmes, and is still the molluscicide of choice [2], since it is effective against most aquatic snails and its activity persists for several months [3]. Due to its high activity at

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all stages of the snail life cycle [4], it has proved to be effective as molluscicidal [5], ovicidal [6], and anthelmintic agent [7]. Besides, niclosamide is introduced as an official drug in many pharmacopoeas [8]. Although certain algae, aquatic plants, are damaged, it does not adversely affect any economically important crop plants and shows no cumulative toxicity on animals [9].

Therefore, the present investigation has been aimed to synthesize new niclosamide Mannich base derivatives 2a–2d and to react them with active nucleophilic phosphacumulene reagents, namely (N-phenyliminovinylidene)- (3a), (2-oxovinylidene)- (3b), and/or (2-thioxovinylidene)triphenylphosphorane (3c). These active phosphacumulene ylides are some of the most important phosphorus reagents, used for the synthesis of heterocyclic phosphorus compounds [10]. A comparative study on the behavior of the *Mannich* bases $2a-2d$ towards the stabilized phosphonium ylides 8a–8f or 12 and the iminophosphorane 14 has been performed, too.

Results and Discussion

The Mannich reaction was carried out in neutral media using the bifunctional compound niclosamide (1) as substrate, formaldehyde and a base as the reagents. The bases employed were dimethyl-, diethylamine, piperidine, and morpholine. The Mannich bases produced 2a–2d were isolated as yellow crystalline solid free bases. Only one product was isolated when either one or two mol-equivalents of the reagents were employed, which proves the presence of only one reactive centre. Absence of the OH absorption band in the IR of 5-chloro-N-(2-chloro-4-nitrophenyl)-2-hydroxy-3-(dimethylaminomethyl)-benzamide (2a) is due to the presence of the intramolecular hydrogen bonding of the hydroxyl hydrogen with the basic nitrogen being a weak band which has been superimposed upon the NH absorption band near $\bar{\nu} = 3420 \text{ cm}^{-1}$. The IR also shows the carbonyl amide at 1671 cm⁻¹. The ¹H NMR spectrum of compound 2a shows bands at $\delta = 2.8$ (2CH₃, s), 4.2 $(CH₂, s)$, 4.5 (NH, s) ppm exchangeable with $D₂O$. The aromatic protons appear at $\delta = 8.2 - 8.9$ (5H, aromatic, m) ppm. Moreover, the OH appears at $\delta = 15$ ppm

Scheme 1

(exchangeable with D_2O). Presence of carbonyl amide group in 2a is also attested by a signal at $\delta = 169.72$ (C=O, NH) in its ¹³C NMR spectrum, and in the mass spectrum m/e is found at 384 (M⁺) (Scheme 1).

When the active phosphacumulene ylides, namely, (N-phenyliminovinylidene)-(3a), $(2-\alpha x)$ (2-oxovinylidene)- (3b), and/or $(2-\alpha x)$ thioxovinylidene)triphenylphosphorane (3c), reacted with niclosamide Mannich bases 2a–2d in dry boiling toluene for 4 h, in case of 3a, 6 h with 3b, and for 8 h with 3c by addition with cyclization, the corresponding phenyliminopyranone 6a, pyranone 6b, and thioxopyranone 6c were formed. The structure of 6-chloro-N-(2-chloro-4-nitrophenyl)-3,4-dihydro-2-(phenylimino)-3-(triphenylphosphoranylidene)-2H-1-benzopyran-8-carboxamide (6a) was elucidated from elemental analysis and spectroscopic data. The ¹H NMR spectrum of 6a shows signals at $\delta = 3.3$ (CH₂, d, due to P atom), 4.2 (NH, s) exchangeable with D_2O and 7.3–8.3 (27H, aromatic, m) ppm. Its IR spectrum shows no OH absorption band which is exhibited by the Mannich bases 2. Absorption bands are shown by compound 6a at $\bar{\nu} = 3419$ (NH), 1674 (C=O, NH), 1610 (P=C) [11], and 1436 (P-aryl) cm⁻¹ [12]. The ³¹P NMR of 6a recordes a signal at δ = 16.82 ppm, which supports the phosphoranylidene structure [13]. In the MS of **6a** the $m/e = 716$ (M⁺) (Scheme 2).

Compounds 6a–6c are equally obtained, irrespective whether one or two molequivalents of the phosphacumulenes 3a–3c are used.

When the Wittig reaction was carried out on pyranone derivative **6b** using p-nitrobenzaldehyde in dry boiling toluene for 8 h, the new exocyclic olefin, 6-chloro-N-(2-chloro-4-nitrophenyl)-3,4-dihydro-3-[(nitrophenyl)methylene]-2-oxo-2H-1-benzopyran-8-carboxamide (7), and triphenylphosphine oxide were obtained. The IR of 7 shows strong absorption bands at $\bar{\nu} = 3419$ (NH), 1739 (C=O, pyran), 1674 (C=O, NH) cm⁻¹, and lacks the presence of a peak arround 1440 (P–C) cm⁻¹. Its ¹H NMR spectrum exhibits signals at $\delta = 4.2$ (CH2, s), 4.6 (NH, s) exchangeable with D₂O. In the MS of 7 the $m/e = 514$ (M⁺) (Scheme 2).

We have also found that the niclosamide *Mannich* bases 2a-2d reacted with the stabilized methylenetriphenylphosphoranes, namely, acetyl- (8a), methoxycarbonyl- (8b), ethoxycarbonyl- (8c), formyl- (8d), or benzoyl-methylenetriphenylphosphorane $(8e)$, in dry boiling toluene for 10 h to give yellow crystalline phosphoranylidenes 11a–11e. Compounds 11a–11e are equally obtained whether one or two mol-equivalents of the Wittig reagents 8 were used with respect to one mol-equivalent of 2. On the other hand, compound 2 reacted with phenylmethylenetriphenylphosphorane (8f) in boiling ethanol for 5 h to give the phosphorane 11f. The IR spectrum of 5-chloro-N-(2-chloro-4-nitrophenyl)-2-hydroxy-3-[3-oxo-2- (triphenylphosphoranylidene)butyl]benzamide (11a) taken as an example reveals the presence of absorption bands at $\bar{v} = 3544$ (OH), 3468 (NH), 1672 (C=O, OCH₃), 1662 (C=O, NH), and 1448 (P-aryl) cm⁻¹. Its ¹H NMR spectrum showed signals at $\delta = 2.03$ (CH₃, s), 3.3 (CH₂, d), 5.08 (NH, s), and 12.3 (OH) ppm exchangeable with D₂O and the aromatic protons appeared at $\delta = 7.2-8.4$ (20H, aromatic, m) ppm. Furthermore signals at $\delta = 169.45$ and 166.56 ppm were observed in the 13 C NMR spectrum which were attributed to the carbonyl group. Moreover, a signal at $\delta = +23.34$ was found in the ³¹P NMR of 11a (Scheme 2).

In the same sense, diphenylmethylenetriphenylphosphorane (12) reacted with niclosamide Mannich bases 2 in ethanol to give the oxaphosphinin 13. The elemental analysis, IR, ${}^{1}H$, ${}^{13}C$ and ${}^{31}P$ NMR data agree with structure 13. A signal at $\delta = +21.66$ was observed in the ³¹P NMR spectrum of compound 13, which is in accordance with the oxaphosphinin structure [14] (Scheme 2).

The reactions of phosphinimines are often analogous to those of phosphonium ylides. But in their activity iminophosphoranes are inferior to phosphin alkylenes [15]. We have found that niclosamide *Mannich* bases $2a-2d$ are inactive against ethoxycarbonyltriphenylphosphinimine (14), even if the reactants were boiled in toluene for a long time, they were recovered practically unchanged.

Biological Evaluation of the Tested Compounds

Materials

Snails: Field collected *B. alexandrina* snails (6–8 mm in diameter) were used to test the molluscicidal potency of the tested chemicals, new Mannich bases and organophosphorus niclosamide derivatives 2a–2d, 6a–6c, 11a–11f, and 13. Snails were obtained from Abu-Rawash, Giza, Egypt and were maintained in the laboratory in glass aerated aquaria, filled with dechlorinated water and kept at 25° C, fed fresh lettuce leaves adlib and left for 45 days to ensure that they were free from infection.

Chemistry of Phosphorus Ylides 245

Chemicals: The tested molluscicides, namely, niclosamide Mannich bases and niclosamide organophosphorus derivatives, were prepared as mentioned in the text.

It is well known that organophosphorus compounds are the most important group of pesticides due to their rapid metabolism, absence of accumulation in the organism of animals, decomposition in the soil, and low chronic toxicity [16]. The use of molluscicides has always been considered to be a major supportive procedure in integrated schistosomiasis control [2]. Among synthetic compounds, niclosamide (the active ingredient of bayluscide) is still the molluscicide of choice, being highly active at all stages of the snail life cycle, effective on the schistosome larvae, and not toxic to humans, domestic animals, and crops [17].

Therefore, the molluscicidal potency of the newly synthesized Mannich bases 2a–2d and organophosphorus niclosamide derivatives 6a–6c, 11a–11f, and 13 against Biomphalaria alexandrina snails, which is the specific intermediate host to Schistosoma mansoni, was studied and compared with bayluscide. The results showed that all the newly synthesized niclosamide derivatives $2a-2d$, $6a-6c$, $11a-$ 11f, and 13 have molluscicidal potency against *B. alexandrina* snails but most of them recorded higher LC_{50} values, except compound 11f which showed a lower value (LC₅₀ of 0.0004 g/dm³) compared to bayluscide (LC₅₀ of 0.0005 g/dm³) [9].

The presence of an OH group in the ortho position of niclosamide (1) is responsible for its toxicity against B. alexandrina snails [18], due to the formation of an intramolecular hydrogen bond with the oxygen of the carbonyl group [18]. This hydrogen bond reduced the polarity of the carbonyl group by decreasing the partial negative charge on the oxygen atom of this group. This in turn increases the chance of bonding with aminoacids of snail proteins most of which are enzymes.

The variable reported molluscicidal activity of the tested newly synthesized compounds could be explained on the basis that all of them still have the reactive

Cmp. no.	LC_5	LC_{10}	LC_{50}	LC_{90}
	g/dm^3	g/dm^3	g/dm^3	g/dm^3
1			0.0005	
2a	0.103	0.0106	0.0301	0.06
2 _b	0.0101	0.0106	0.0409	0.10
2c	0.009	0.0108	0.019	0.0293
2d	0.005	0.0101	0.030	0.08
6a	0.00098	0.0021	0.009	0.0303
6b	0.0022	0.0044	0.0077	0.021
6c	0.00097	0.0012	0.0041	0.007
11a	0.002	0.0036	0.0052	0.0076
11 _b	0.0104	0.0202	0.030	0.05
11c	0.006	0.0109	0.0208	0.0404
11e	0.0018	0.0021	0.0034	0.0056
11f	0.0009	0.00012	0.0004	0.0006
11d	0.0024	0.003	0.007	0.0098
13	0.0009	0.0009	0.0019	0.004

Table 1. Molluscicidal potency of the tested compounds against *B. alexandrina* snails

 LC_X : The concentration at which $X\%$ of the populations died

groups of the niclosamide nucleus with different chemically reactive phosphorus moieties added, most of them are organophosphorus groups.

On the other hand, the biochemical investigation has been performed on snail metabolism. The most potent organophosphorus molluscicides 11f, 11e, 13, and 6c, showing LC_{50} values between 0.0004–0.0041 g/dm³ were selected for this study. We found that these four organophosphours niclosamide derivatives have an inhibitory effect on the most important glycolytic enzymes in tissue homogenates of B. alexandrina snails, as hexokinase (HK), pyruvate kinase (PK), and phosphoglucoisomerase (PGI). Although bayluscide showed a slightly higher inhibitory effect on hexokinase (HK) and glucose phosphate isomerase (PGI) compared to the organophosphorus derivatives 11f and 13, the four niclosamide derivatives 6c, 11e, 11f, and 13 showed a remarkably higher inhibition rate of pyruvate kinase. Pyruvate kinase is an enzyme of critical importance for schistosome parasites, as homolactate fermenter within snail tissues [19].

Inhibition of hexokinase, pyruvate kinase, and glucose phosphate isomerase with the four tested compounds 6c, 11e, 11f, and 13 and of hexokinase together with glucose phosphate isomerase only with bayluscide could be effective in decreasing the compatibility of *B. alexandrina* snails and hence *S. mansoni* infection.

It was found that the sublethal and less pollutant concentrations, LC_5 (1/10 of LC_{50}) were able to affect the compatibility of B. alexandrina to parasitic infection.

Snail group	Enzymes	HK	PK	PGI	
Control 1-Bayluscide treated	Mean \pm S.D. Mean \pm S.D. P< $%$ change	9.8 ± 2.7 3.9 ± 0.58 0.01 59.4	1.8 ± 0.62 1.51 ± 0.52 n.s. 11.11	1.47 ± 0.48 0.89 ± 0.21 0.01 39.17	
6c -Treated	Mean \pm S.D.	2.6 ± 0.82	0.98 ± 0.138	1.27 ± 0.32	
	P<	0.001	0.01	n.s.	
	$%$ change	72.9	44.4	13.6	
11e-Treated	Mean \pm S.D.	8.0 ± 0.49	0.46 ± 0.08	1.29 ± 0.19	
	P<	0.05	0.001	n.s.	
	$%$ change	16.6	72.2	12.24	
11f-Treated	Mean \pm S.D.	3.7 ± 1.6	0.39 ± 0.29	1.19 ± 0.38	
	P<	0.01	0.01	n.s.	
	$%$ change	61.4	77.8	19.0	
13-Treated	Mean \pm S.D.	9.4 ± 0.32	0.304 ± 0.1	1.17 ± 0.42	
	P<	n.s.	0.001	n.s.	
	$%$ change	2.08	83.3	20.4	

Table 2. Levels of hexokinase (HK), pyruvate kinase (PK), and glucose phosphate isomerase (PGI) in field collected, control and molluscicide-treated snails

– Values are means \pm S.D. of four independent experiments

 $- P < 0.05 \equiv$ significant, $P < 0.01 \equiv$ highly significant, n.s. $=$ no significance

– Enzymatic activities are expressed as mol/min/g tissue

 $-$ S.D. $=$ Standard deviation

 $-$ % change = Percent change from control group

Snails treated with LC_5 of the four tested compounds 6c, 11e, 11f, and 13 were metabolically disturbed and render less suitable for the development of the parasite (S. mansoni larvae) within snails tissue compared to bayluscide (1). This inspires more hope for their application in the intervention of schistosomiasis, especially, when used at their sublethal concentrations (LC_5) .

Conclusion

The reaction of the bifunctional Mannich base derivatives 2a–2d with phosphacumulene ylides 3a–3c represents a new approach to the construction of new heterocycles. These phosphonium ylides react only with the phenolic OH group rather than the NH group of the *Mannich* bases $2a-2d$, to give first the phosphonium ylides 4, which are transformed into 5 by nucleophilic substitution of the dialkylamine anion. Elimination of the amine anion from 5 results in the formation of phenylimino-, oxo-, and thioxophosphoranylidene benzopyran carboxamide derivatives 6a–6c. On the other hand, transylidation of niclosamide Mannich base derivatives 2a–2d, using the stabilized methylenetriphenylphosphoranes 8a–8f or 12, results in the expulsion of the amine, with the formation of the unstable quinone-methylene 9. The latter reacts with the phosphoranes 8 by nucleophilic addition to the methylene group giving the dipolar adduct 10. The R moiety in 10, which is electron withdrawing in nature, would stabilize formation of the new alkylated phosphoranylidene benzamide derivatives 11 *via* migration of the α -proton to the electron rich centre of the molecule. Moreover, the quinone-methylene 9 affords directly the oxaphosphinin 13, in case, when diphenylmethylenetriphenylphosphorane (12) reacts with niclosamide Mannich bases 2. No reaction was observed when the iminophosphorane 14 was allowed to react with niclosamide Mannich bases 2 under different reaction conditions.

Experimental

All melting points are uncorrected. The solvents were dried and distilled. Reactions were carried out under nitrogen atmosphere. Elemental analyses were carried out at the ''Microanalysis Department'', National Research Centre; their results were in agreement with the calculated values. The IR spectra were measured in KBr on a Perkin-Elmer infracord Spectrometer Model 157 (Grating). The ¹H and ¹³C NMR spectra were recorded on a Varian Spectrometer at 90 (22.5) MHz, using TMS as an internal reference. ³¹P NMR spectra were run, relative to external H_3PO_4 (85%), with a Varian FT-80 Spectrometer at 36.5 MHz, mass Spectra were obtained on a Varian MATCH-4B instrument.

Mannich Reaction on Niclosamide (5-Chloro-N-(2-chloro-4-nitrophenyl)-2 hydroxybenzamide; 1)

General Procedure: An aqueous solution of formaldehyde (40%, 0.01 mol) was added dropwise with stirring to niclosamide (1) [20] and the amine (0.011 mol) in about 40 cm³ of ethanol while maintaining the temperature below 10° C. The reaction mixture was then boiled under reflux for 2h and left overnight at room temperature. After removing the volatile materials under reduced pressure, the product was isolated and recrystallized from ethanol. When the above described procedure was

Cpd. no.	Mol. Wt/g mol ⁻¹	Formula	Mp [°] C	Yield/%	Solvent for crystallization
2a	384	$C_{16}H_{15}N_3O_4Cl_2$	228	92%	Ethanol
2 _b	412	$C_{18}H_{19}N_3O_4Cl_2$	340	90%	Ethanol
2c	424	$C_{19}H_{19}N_3O_4Cl_2$	232	87%	Ethanol
2d	426	$C_{18}H_{17}N_3O_5Cl_2$	300	95%	Ethanol
6a	716	$C_{40}H_{28}N_3O_4PCl_2$	244	87%	Chloroform/Pet.ether
6b	641	$C_{34}H_{23}N_2O_5PCl_2$	264	85%	Ethyl acetate/Pet.ether
6c	657	$C_{34}H_{23}N_2O_4SPCl_2$	252	83%	Chloroform/Pet.ether
7	514	$C_{23}H_{13}N_3O_7Cl_2$	214	38%	Chloroform/Light pet.
11a	657	$C_{35}H_{27}N_2O_5PCl_2$	220	79%	Ethyl acetate/ n -Hexane
11 _b	673	$C_{35}H_{27}N_2O_6PCl_2$	228	80%	Acetone/ n -hexane
11c	657	$C_{36}H_{29}N_2O_6PCl_2$	188	79%	Benzene/Pet.ether
11d	643	$C_{34}H_{25}N_2O_5PCl_2$	245	78%	Methylene chloride/ n -Hexane
11e	719	$C_{40}H_{29}N_2O_5PCl_2$	186	82%	Chloroform/ n -Hexane
11f	703	$C_{40}H_{29}N_2O_4PCl_2$	215	76%	Ethyl acetate/ <i>n</i> -Hexane
13	780	$C_{46}H_{34}N_2O_4PCl_2$	205	74%	Methylene chloride/ n -Hexane

Table 3. Physical and analytical data for niclosamide Mannich bases 2a–2d, phenyliminopyranone 6a, pyranone 6b, thioxopyranone 6c, exocyclic olefin 7, phosphoranylidenes 11a–11f, and oxaphosphinin 13

performed using two mol-equivalents of both the base and aqueous formaldehyde no change in the nature of the products was observed and the corresponding niclosamide *Mannich* bases 2a–2d were obtained, respectively.

The Reaction of Niclosamide Mannich Base $2c$ with Phosphacumulenes $3a-3c$ Preparation of Phenyliminopyranone 4a, Pyranone 4b, and Thioxopyranone 4c

General Procedure: To a solution of niclosamide Mannich base 2c (0.01 mol) in 20 cm³ of dry toluene, was added a solution of (N-phenyliminovinylidene)- $(3a)$ [21], (2-oxovinylidene)- $(3b)$ [22], and (2thioxovinylidene)triphenylphosphorane (3c) [22] (0.011 mol) in 30 cm³ of dry toluene. The reaction mixture was refluxed for 4 h when 3a was used, 6 h in case of 3b, and for 8 h with 3c. After the solvent had been distilled off under reduced pressure, the residue was crystallized from the appropriate solvent. Yields, analytical and spectroscopic data are presented in Tables 3 and 4.

When the reaction was performed using one mol-equivalent of niclosamide Mannich base derivative 2d and the phosphacumulenes 3a–3c, the same phenyliminopyranone 6a, pyranone 6b, and thioxopyranone 6c were obtained irrespective of the amine used.

When the reaction was repeated using one mol-equivalent of niclosamide *Mannich* base 2c and two mol-equivalents of the phosphacumulenes 3a–3c, the same products 6a–6c were isolated, too.

The Reaction of Pyranone 6b with p-Nitrobenzaldehyde

A mixture of the pyranone derivative 6b (0.64 g, 0.001 mol) and p-nitrobenzaldehyde (0.17 g, 0.0011 mol) in dry toluene (20 cm^3) was refluxed for 8 h. Toluene was distilled off and the residue was crystallized from benzene to give the exocyclic olefin 7, 6-chloro-N-(2-chloro-4-nitrophenyl)-3,4-dihydro-3-[(nitrophenyl)methylene]-2-oxo-2H-1-benzopyran-8-carboxamide (7) as pale greenish crystals.

The benzene filtrate yielded upon concentration and addition of n -hexane colourless crystals of triphenylphosphine oxide, mp and mixed mp 151° C [23] (62%).

Cpd. no	IR: $\bar{\nu}/\text{cm}^{-1}$	NMR: δ /ppm			
		¹ H NMR	$31P$ NMR	13 C NMR	
2 _b	3421 (NH), 1670 $(C=O, amide)$	1.3 (t, 3H, CH_2CH_3) 3.1 (s, 2H, $CH2$) 4.3 (q, 2H, \underline{CH}_2CH_3) 4.5 (s, 1H, NH) $7.5-8.3$ (m, 5H, Ar) 15 (s, 1H, OH)		168.54 $(C=O, amide)$	
2c	3422 (NH), 1655 $(C=O, amide)$	3.8 (s, 2H, $CH2$) 4.1 (s, 1H, NH) $7.1-8.7$ (m, 15H, Ar) 8.3 (s, 1H, OH)			
2d	3423 (NH), 1671 $(C=O, amide)$	3.5 (s, 2H, $CH2$) 4.4 (s, 1H, NH) $7.2 - 8.2$ (m, 13H, Ar) 8.6 (s, 1H, OH)			
6b	3429 (NH), 1731 (C=O, pyran), 1645 (C=O, amide), 1439 (P-aryl)	3.3 (d, 2H, $CH2$) 5.3 (s, 1H, NH) $7.3-8.4$ (m, 20H, Ar)	21.26	165 (C=O, amide) 185 (C=O, pyran)	
6c	3427 (NH), 1635 (C=O, amide), 1439 (P-aryl), 1240 $(C=S)$	3.21 (d, 2H, $CH2$) 3.9 (s, 1H, NH) $7.2 - 8.4$ (m, 20H, Ar)	25.82		
11b	3456 (OH), 3118 (NH), 1674 (C=O, ester), 1662 $(C=O, amide),$ 1447 (P-aryl)	2.7 (s, $3H$, OCH ₃) 3.8 (d, 2H, $CH2$) 4.5 (s, 1H, NH) $7.0-8.4$ (m, 20H, Ar) 12.1 (s, 1H, OH)	21.55	165.72 163.52 $(C=O)$	
11c	3446 (OH), 2942 (NH), 1671 (C=O, ester), 1663 $(C=O, amide), 1448$ (P-aryl)	2.4 (d, 2H, $CH2$) 2.8 (t, 3H, CH_2CH_3) 3.82 (q, 2H, \underline{CH}_2CH_3) $7.1-8.3$ (m, 20H, Ar) 11.28 (s, 1H, OH)	20.66		
11d	3484 (OH), 3120 (NH), 1733 (CHO), 1661 (C=O, amide), 1447 (P-aryl)	2.5 (d, 2H, $CH2$) 4.3 (s, 1H, NH) 7.3-8.4 (m, 20H, Ar) 12 (s, 1H, OH)	22.55		
11e	3566 (OH), 3488 (NH), 1673 (C=O, ester), 1660 $(C=O, NH)$, 1434 (P-aryl)	2.33 (s, 1H, NH) 3.81 (d, 2H, $CH2$) $7.3 - 8.2$ (m, 25H, Ar) 11.9 (s, 1H, OH)	22.23	183.25 168.62 $(C=O)$	
11f	3421 (OH), 2364 (NH), 1664 (C=O), 1436 (P-aryl)	2.8 (d, 2H, $CH2$) 4.7 (s, 1H, NH) 8.0 (s, 1H, OH) $7.2 - 8.1$ (m, 25H, Ar)	22.03	169.74 $(C=0)$	
13	3558 (NH), 1663 (C=O), 1447 (P-aryl)	2.4 (d, 2H, $CH2$) 4.1 (s, 1H, NH) $7.0-8.0$ (m, 30H, Ar)	21.66	169.73 $(C=0)$	

Table 4. Spectroscopic data (IR & NMR) for compounds 2b–2d, 6b–6c, 11b–11f, and 13

The Reaction of Niclosamide Mannich Base 2c with the Stabilized Phosphonium Ylides 8a–8e. Preparation of the Phosphoranylidenes 11a–11e

General Procedure: To a solution of niclosamide Mannich base $2c$ (0.01 mol) in 20 cm³ of dry toluene, was added a solution of the methylenetriphenylphosphoranes $8a-8e$ [24–26] (0.01 mol) in 30 cm³ of dry toluene and the reaction mixture was refluxed for 10 h. After the solvent was distilled off, the residue was crystallized from the appropriate solvents. Yields, analytical and spectroscopic data are presented in Tables 3 and 4.

The Reaction of Niclosamide Mannich Base $2c$ with Phenylmethylenetriphenylphosphorane (8f) and Diphenylmethylenetriphenylphosphorane (12). Synthesis of the Phosphoranylidene 11f and Oxaphosphinin 13

A solution of the Wittig reagents 8f [27] or 12 [28] (0.01 mol) in 20 cm³ of absolute ethanol was treated with 20 cm^3 of a $0.01 M$ solution of sodium ethoxide in ethanol with continuous stirring, where the orange colour of the phosphorane appeared immediately. The phosphorane solution was then added to a solution of the *Mannich* base 2c in 30 cm^3 of absolute ethanol and the reaction mixture was stirred and boiled for 2h at 25° C. Ethanol was distilled off under reduced pressure and the remaining precipitate was extracted with dry benzene. After the benzene extract had been concentrated, the phosphoranylidene 11f and the oxaphosphinin 13 were precipitated. Yields, analytical and spectroscopic data are presented in Tables 3 and 4.

Attempted Reaction of Niclosamide Mannich Base 2c with Ethoxycarbonyltriphenylphosphinimine (14)

No reaction was observed between the niclosamide Mannich base 2 and the phosphinimine 14 [29], even if the reactants were boiled in THF or toluene for a long time, they were recovered unchanged.

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Chemistry of Phosphorus Ylides 251

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